## Structural changes in the Mn<sub>4</sub>Ca cluster and the mechanism of photosynthetic water splitting

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Photosynthetic water oxidation, where water is oxidized to dioxygen, is a fundamental chemical reaction that sustains the biosphere. This reaction is catalyzed by a Mn<sub>4</sub>Ca complex in the photosystem II (PS II) oxygen-evolving complex (OEC): a multiprotein assembly embedded in the thylakoid membranes of green plants, cyanobacteria, and algae. The mechanism of photosynthetic water oxidation by the Mn<sub>4</sub>Ca cluster in photosystem II is the subject of much debate, although lacking structural characterization of the catalytic intermediates. Biosynthetically exchanged Ca/Sr-PS II preparations and x-ray spectroscopy, including extended x-ray absorption fine structure (EXAFS), allowed us to monitor Mn-Mn and Ca(Sr)-Mn distances in the four intermediate S states, So through S3, of the catalytic cycle that couples the one-electron photochemistry occurring at the PS II reaction center with the four-electron water-oxidation chemistry taking place at the Mn<sub>4</sub>Ca(Sr) cluster. We have detected significant changes in the structure of the complex, especially in the Mn-Mn and Ca(Sr)-Mn distances, on the S2-to-S3 and S3-to-S0 transitions. These results implicate the involvement of at least one common bridging oxygen atom between the Mn-Mn and Mn-Ca(Sr) atoms in the O-O bond formation. Because PS II cannot advance beyond the S2 state in preparations that lack Ca(Sr), these results show that Ca(Sr) is one of the critical components in the mechanism of the enzyme. The results also show that Ca is not just a spectator atom involved in providing a structural framework, but is actively involved in the mechanism of water oxidation and represents a rare example of a catalytically active Ca cofactor.

manganese enzyme | oxygen evolution | photosynthesis | photosystem II | x-ray spectroscopy

Photosynthetic oxidation of water to dioxygen is catalyzed by a Mn<sub>4</sub>Ca cluster in the oxygen-evolving complex (OEC) of photosystem II (PS II) (1). PS II is a multiprotein assembly embedded in the thylakoid membranes of green plants, cyanobacteria, and algae. The structure and function of the Mn<sub>4</sub>Ca cluster, the heart of the OEC, has been under active investigation by many spectroscopic techniques such as EPR/electron-nuclear double resonance (ENDOR) (2-4), x-ray spectroscopy (5, 6), and FTIR (7, 8). X-ray diffraction (XRD) studies at 3.0–3.8 Å resolution have located the Mn<sub>4</sub>Ca cluster in the electron-density map and confirmed the presence of Ca in the cluster (9, 10). Before the XRD studies, the presence of Ca in the OEC was detected by using Ca and Sr x-ray absorption spectroscopy (XAS) studies (11–15). Mn, Ca, and Sr extended x-ray absorption fine structure (EXAFS) studies of PS II frozen solutions have provided accurate distances (≈0.02 Å) and information on the numbers of Mn-Mn, Mn-Ca and Mn/Ca-ligand vectors in the Mn<sub>4</sub>Ca cluster (6, 16, 17). XAS experiments require a significantly lower x-ray dose than XRD measurements. The onset of radiation damage can be precisely determined and controlled by monitoring the Mn K-edge position, thus allowing us to collect data from the intact Mn<sub>4</sub>Ca cluster of PS II. Recently, a high-resolution Mn<sub>4</sub>Ca structural model was determined by using a polarized Mn EXAFS study on PS II single crystals in the  $S_1$  state (18).

Of all the cations that can competitively replace Ca in PS II, only Sr is capable of supporting  $O_2$  evolution, a feature that has been attributed to the similar electronegativities of Ca and Sr and, hence, to their similar Lewis acidities and  $pK_a$  values of water ligands (19, 20). Substituting Sr for Ca alters the EPR properties of the  $S_2$  state, increasing the fraction of PS II that exhibits a g  $\approx$  4.1 or g  $\approx$  5.25 EPR signal and altering the appearance of the g = 2 multiline EPR signal (19, 21). The presence of Sr also shifts a low-frequency  $S_2$  state vibrational mode that has been assigned to a Mn–O–Mn structural unit (22) and alters several carboxylate-stretching modes of the  $S_2-S_1$  FTIR difference spectrum (23–26). This demonstrates that Ca substitution for Sr perturbs the structure of  $Mn_4$  cluster only slightly, making Sr a relevant spectroscopic probe.

This x-ray absorption study determines the changes in the Mn–Mn and Ca(Sr)–Mn distances in the four intermediate S states, S<sub>0</sub> through S<sub>3</sub>, of the catalytic cycle. This became possible by the use of unique PS II preparations from the thermophilic cyanobacterium, *Thermosynechococcus elongatus*, with Ca biosynthetically replaced by Sr in an isomorphic structure (27). The advantages over Ca/Sr exchange induced by biochemical procedures are: (*i*) an enzyme fully competent in O<sub>2</sub> evolution, (*ii*) a 1:1 Sr-to-PS II ratio monitored by Sr quantitation, and (*iii*) Sr K-edge EXAFS measurements that result in less x-ray damage and are preferred for experimental reasons to those at lower-energy Ca K-edge (13–15).

## Results

EPR Spectroscopic Quantitation of S State Composition. Protein samples in the  $S_1$  state were advanced to primarily  $S_2$ ,  $S_3$ , and  $S_0$  states by giving one, two, and three laser flashes followed by immediate freezing in liquid nitrogen (28). Starting from a dark-adapted sample, the  $S_2$  state multiline EPR signal is maximal after one flash and oscillates with a period of four as a function of flash number; thus, the  $S_2$  state multiline EPR signal can be used to characterize the S state distribution of samples given zero, one, two, three, four, five, or six flashes. Supporting information (SI) Fig. 5.4 shows typical EPR spectra obtained after a given flash number. A deep period-four oscillation is observed with maxima on the first and fifth flashes, indicating that there is very little dephasing of S state advancement with flashes and that the initial S state population is close

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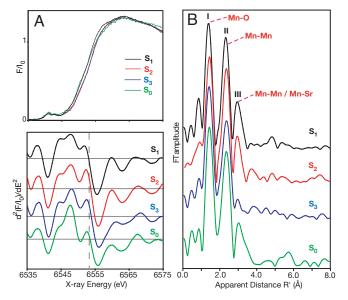


Fig. 1. Mn XAS of Sr-PS II from *T. elongatus*. (A) Mn K-edge XANES spectra (Upper) and the corresponding second derivatives of the XANES spectra (Lower) in the  $S_1$ ,  $S_2$ ,  $S_3$ , and  $S_0$  states. The inflection point of the edges and the shape of the spectra are clearly different between the S states. (B) Fourier transforms of k3-weighted Mn EXAFS (SI Fig. 6) of Sr-PS II in the different S states (S<sub>1</sub>, S<sub>2</sub>, S<sub>3</sub>, and S<sub>0</sub>). The detailed fit parameters of the Mn EXAFS spectra are included in Table 1 and SI Tables 5 and 6.

to 100% S<sub>1</sub>Y<sub>D</sub><sup>ox</sup>. The S state composition of each sample was determined by EPR (SI Fig. 5 and SI Tables 3 and 4), and these data were used to generate the x-ray spectra of the pure S states.

X-ray Absorption Spectroscopy at the Mn K-Edge. Mn x-ray nearedge spectra (XANES) and EXAFS data were collected for all S state samples, ensuring the integrity of the samples and the absence of radiation damage to the structure of the Mn<sub>4</sub> core (Fig. 1). The resulting Mn spectra were similar to those from the same S states in Ca-containing PS II protein complexes isolated from spinach (28, 29) or the thermophilic cyanobacterium (18, 30). The edge shift that is apparent between the  $S_1$  and  $S_2$  spectra (Fig. 1A) indicates that Mn oxidation occurs during the  $S_1$ -to- $S_2$ transition. However, it is less clear that there is a shift between the S<sub>2</sub> and S<sub>3</sub> spectra, which suggests that Mn is not oxidized as the OEC proceeds from the  $S_2$  to  $S_3$  state (28). The edge position shifts to lower energy between the S<sub>3</sub> and S<sub>0</sub> spectra, as expected for  $S_3 \rightarrow [S_4] \rightarrow S_0$  transition.

In the Fourier transform (FT) (Fig. 1B) of the Mn EXAFS data (SI Fig. 6), FT peak I is characteristic of the 1.8-Å Mn-oxo bridging ligand distances, FT peak II is best fit to two to three di- $\mu$ -oxo-bridged Mn–Mn interactions at  $\approx$ 2.7 Å (Table 1), and FT peak III is from one mono-μ-oxo-bridged Mn–Mn and from Mn-Sr interactions. The presence of these three Fourier peaks is diagnostic for an intact bridged multinuclear Mn<sub>4</sub>Ca(Sr) complex. The Mn<sub>4</sub>Sr and Mn<sub>4</sub>Ca clusters are essentially identical in terms of the structure of Mn<sub>4</sub>-core; this is not surprising, because these clusters exhibit only small differences in their kinetic and spectroscopic properties.

Mn EXAFS of the S<sub>1</sub>, S<sub>2</sub>, and S<sub>0</sub> states are similar; however, as Fig. 1B shows, the S<sub>3</sub> state spectrum differs from those of the other S states. In the S<sub>3</sub> state the intensity of FT peak II decreases and peak III is significantly attenuated. This is best explained by changes in Mn-Mn and Mn-Sr distances and thus by structural changes in the OEC on the  $S_2$  to  $S_3$  and  $S_3 \rightarrow [S_4] \rightarrow S_0$  transitions (Table 1 and SI Tables 5 and 6). The low intensity of FT peak III (there are 4 Mn/1 Sr) prevents reliable analysis of Mn-Sr interaction from the "Mn" point of view; however, as demonstrated below, the Sr EXAFS technique can clearly resolve the Sr-Mn interaction from the "Sr" point of view in all of the S

X-ray Absorption Spectroscopy at the Sr K-Edge. The Sr K-edge XANES of Sr-PS II, which corresponds to dipole-allowed transitions from the 1s core level to molecular orbitals with predominant p character, contains information about the valence and coordination environment of the absorbing atom. The Sr XANES spectra of Sr-PS II show that there is a pronounced difference between the S state spectra and the spectrum of inactive Sr-PS II after the Mn<sub>4</sub>Sr cluster is disrupted by hydroxylamine (HYD) (Fig. 24). This shows that in active PS II the Sr first coordination environment is different from that of free Sr<sup>2+</sup> ion. Moreover, there are differences in the Sr XANES in the  $S_1$ ,  $S_2$ ,  $S_3$ , and  $S_0$  states, indicating that Sr(Ca) is an integral part of the cluster. The Sr XANES is sensitive to changes in the electronic structure caused by changes in the oxidation state and geometry of the cluster as it cycles through the S states.

The structural changes of the S states are even more obvious in the Fourier transforms (FTs) (Fig. 2B) of the EXAFS spectra (SI Fig. 7) of Sr-PS II in the  $S_1$ ,  $S_2$ ,  $S_3$ , and  $S_0$  states and the

Table 1. One- and two-shell simulations of Mn EXAFS Fourier peak II from the S1, S2, S3, and S₀ states

Fit	Sample	Shell	R, Å	N	$\sigma^2$ ( $ imes$ 10 $^3$ ), Å $^2$	$\Phi$ ( $ imes$ 10 $^3$ )	$arepsilon^2$ ( $ imes$ 10 $^5$ )
				One sh	ell		
1	S <sub>1</sub>	Mn–Mn	2.73	1.1	1.7	0.20	0.10
2	$S_2$	Mn–Mn	2.74	1.1	1.6	0.22	0.10
3	$S_3$	Mn–Mn	2.76	1.4	3.3	0.31	0.15
4	$S_0$	Mn–Mn	2.73	1.3	2.0	0.30	0.16
				Two she	ells		
5	S <sub>3</sub>	Mn–Mn	2.75	1.1	1.0*	0.23	0.11
		Mn–Mn	2.88	0.5	1.0*		
6	$S_0$	Mn–Mn	2.73	1.1	1.0*	0.28	0.14
		Mn–Mn	2.85	0.3	1.0*		

R is the Mn-Mn distance.  $S_0^2$ , an amplitude reduction factor (see SI Text, Eq. 1), was set to 0.85. Range-extended EXAFS methodology can distinguish two Mn–Mn distances at ≈2.7 and ≈2.8 Å contributing to peak II in the S<sub>1</sub> and  $S_2$  states (16, 17). Peak II in the  $S_1$  and  $S_2$  states can only be fit to one shell of Mn-Mn interactions at the resolution of conventional EXAFS. However, a two-shell fit is favored for the S<sub>0</sub> and S<sub>3</sub> states as indicated by an increased Debye–Waller factor for the one-shell fit.  $\Phi$  and  $\varepsilon^2$  are the goodness-of-fit parameters (see *SI Text*). \*Parameter was fixed.

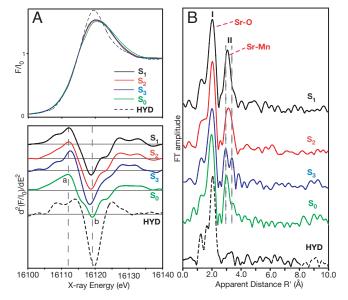


Fig. 2. Sr XAS of Sr-PS II from T. elongatus. (A) Sr K-edge XANES spectra (Upper) and the corresponding second derivatives of the XANES spectra (Lower) in the S<sub>1</sub>, S<sub>2</sub>, S<sub>3</sub>, and S<sub>0</sub> states and an inactive control sample (HYD). The inflection point of the edges and the shape of the spectra are clearly different between the control and the intermediate S states. There are small but distinct differences in the S state spectra that are easier to see in the second derivatives (Lower). The two vertical dashed lines indicate the clear systematic differences in the Sr K-edge spectra between the S states; the feature labeled a shifts to higher energy and feature b shifts to lower energy as the OEC advances from  $S_0$  through  $S_3$  states. Other small differences are also seen in the second derivatives. (B) Fourier transforms of  $k^3$ -weighted Sr EXAFS (SI Fig. 7) in the different S states ( $S_1$ ,  $S_2$ ,  $S_3$ , and  $S_0$ ), and an *inactive* control sample prepared by hydroxylamine treatment of Sr-PS II. The dominant Fourier peak I is due to ligating oxygens in the first coordination sphere to Sr. Peak II is best fit by four Sr–Mn interactions; two short distances at  $\approx$ 3.5 and two longer Sr–Mn distances at ≈4.0 Å. FT peak II from Sr–Mn depends on the particular S state and shows that structure of cluster changes as the OEC advances through the S states, with a significant change occurring between the critical S2-to-S3 transition, suggesting that the cluster is flexible. The control sample shows only the FT peak from Sr-O backscattering, because the cluster is disrupted and the  $Sr-Mn interactions \, are \, lost. \, The \, detailed \, fit \, parameters \, of \, the \, Sr \, EXAFS \, spectra \,$ are included in Table 2 and SI Tables 7-9.

inactive control sample (HYD). The two major Fourier peaks, I and II, represent the radial distribution of the x-ray photoelectron backscattering from the atoms surrounding Sr. Fourier peak I is best simulated by one shell of seven to eight oxygen atoms at  $\approx\!2.5$  Å (SI Table 7). Fourier peak II consists of at least two different Sr–Mn interactions at  $\approx\!3.5$  Å and  $\approx\!4.0$  Å in all S states (Table 2 and SI Tables 8 and 9). This Fourier peak is absent in the inactive control sample (HYD), because there are no Sr–Mn interactions in this sample. Relative to the S1 state sample, FT peak II is broadened in S2 and split in the S3 and S0 states; these changes are significant.

## Discussion

Position of Sr Atom in the OEC in the  $S_1$  State. Previously, the Ca EXAFS of the native PS II (14) and Sr EXAFS of Sr-reactivated PS II membranes (13) indicated proximity of the Ca at 3.4 Å and Sr at 3.5 Å to the Mn cluster in the  $S_1$  dark-stable state of the OEC. The present study unambiguously demonstrates that Sr is proximate to the OEC in all S states and that significant changes occur in the Sr(Ca)–Mn interactions as the enzyme proceeds through the catalytic cycle, as seen in the FTs in Fig. 2B and Table 2. The Sr–Mn distances in the recently reported first synthetic heteronuclear Mn/Sr complex (31), where Mn and Sr are bridged by an oxo group, are similar to those observed in PS

II. Therefore, it is likely that the Ca/Sr atom in PS II is bridged by a  $\mu$ -oxo group to Mn atoms.

The best fits (Table 2) to the Sr EXAFS data (Fig. 2B) in the  $S_0$  through  $S_3$  states are obtained for Sr–Mn distances at  $\approx 3.5$  Å (short) and  $\approx 4.0$  Å (long). The best fits for the  $S_1$  state are for 3:1, 2:2, and 2:1 short-to-long Sr–Mn ratio of vectors; however, the 3:1 fit is better than the others by 25–28% (Table 2 and SI Fig. 8). The best fit for the  $S_2$  state is for a 3:1 or 2:2 short-to-long Sr–Mn ratio of vectors, with the fit for 2:2 being better by 21%. For the  $S_3$  and  $S_0$  states, a ratio of 2:2 provides the best fit (Table 2 and SI Fig. 8). This information is used below to model the position of Sr and the important changes in the short and long Sr–Mn distances during the S state transitions.

To model the position of the Sr atom in the  $S_1$  state we combined information from the EXAFS fits (Table 2) and from polarized EXAFS measurements of Ca-containing PS II single crystals (18), from which three high-resolution Mn<sub>4</sub>Ca structural models (I, II, and III) have been proposed (SI Fig. 9). For the three best fits (Table 2), there are three or two Sr-Mn vectors at  $\approx$ 3.5 Å and one or two Sr-Mn vectors at  $\approx$ 4.0 Å. For all three cases the Sr atom is displaced <1.2 Å from the position of Ca in the models, and is directly connected to the Mn core through a bridging oxygen atom. The Ca- and Sr-PS II are slightly different in their kinetic and spectroscopic properties (27), which may reflect differences in the interaction of Ca and Sr with the Mn atoms. Previous results from Ca EXAFS of plant PS II have shown that there are only two Mn-Ca distances (14) at <4 A, compared with the four we have observed in Sr-PS II; it is possible that the two longer interactions are at >4 Å or were not discernible at the signal-to-noise ratio of the Ca EXAFS data (14). There is also evidence that Ca protects two of the four Mn atoms from reductants, suggesting a closer interaction between Ca and two of the four Mn atoms in the cluster (32). In Fig. 3A we use model II (from ref. 18) to illustrate the best possible position of the Sr atom (fit 1 in Table 2; 3:1 ratio) in the S<sub>1</sub> state. The experimental FT from the S<sub>1</sub> state of Sr-PS II is comparable with the FT calculated for the structure shown in Fig. 3A. Models I and III from Yano et al. (18) were also considered. The positions of Mn atoms in model III from Yano et al. (18) are identical to those in model II, resulting in the same Sr positions relative to the Mn<sub>4</sub> core (data not shown). The Sr/Ca atom in model III is  $\approx$ 4 Å away from the  $\mu_3$ -oxo-bridged oxygen, requiring  $\mu$ -oxo bridges between Sr and Mn to explain the presence of short distances. Range-extended Mn EXAFS (17) results from oriented PS II membranes show that model I is less probable than models II or III.

Structural Changes in the OEC. Previous Mn EXAFS experiments demonstrated the absence of major changes in Mn–Mn distances of the Mn<sub>4</sub> core during the  $S_1$ -to- $S_2$  transition. Our present Mn and Sr EXAFS data (Figs. 1B and 2B and Tables 1 and 2) also show that there are no significant changes in the Mn–Mn and Sr–Mn distances during this transition. It is possible that the ratio of short:long Sr–Mn vectors changes from 3:1 ( $S_1$ ) to 2:2 ( $S_2$ ), on the basis of the quality of the fits (Table 2), indicating that one Sr–Mn distance changes from  $\approx$ 3.5 to  $\approx$ 4.0 Å during this transition. However, we cannot rule out the presence of the 2:2 ratio in the  $S_1$  state or the 3:1 ratio in the  $S_2$  state, because of the uncertainties in EXAFS fitting procedures.

A one-shell fit of peak II in the Mn EXAFS of the  $S_3$  state (Table 1) shows an elongation of the Mn–Mn distance and an increased Debye–Waller factor compared with the  $S_1$  and  $S_2$  states. Introduction of the second Mn–Mn subshell improves the fit quality and resolves two distances (2.75 Å and 2.88 Å) among the di- $\mu$ -oxo-bridged Mn–Mn moieties. The low intensity of the Mn EXAFS FT peak III prevents reliable analysis of the changes in the mono- $\mu$ -oxo-bridged Mn–Mn and Mn–Sr interactions  $\approx 3.2–3.5$  Å.

Table 2. Two-shell simulations of Sr EXAFS Fourier peak II (Sr-Mn distances) from the S1, S2, S3, and S0 states

Fit	S state	Shell	R, Å	N* (Sr-Mn interactions)	$\sigma$ $^{2}$ ( $ imes$ 10 $^{3}$ ), Å $^{2}$	$\Phi$ ( $ imes$ 10³)	$arepsilon$ $^2$ ( $ imes$ 10 $^5$ )
1	S <sub>1</sub>	Sr–Mn	3.53	3	7.0	0.45	0.18
		Sr–Mn	4.02	1	7.0		
2	S <sub>1</sub>	Sr–Mn	3.52	2	5.5	0.62	0.25
		Sr–Mn	3.99	2	5.5		
3	$S_1$	Sr–Mn	3.52	2	4.6	0.58	0.24
		Sr–Mn	3.99	1	4.6		
1	$S_2$	Sr–Mn	3.51	2	7.7	0.37	0.15
		Sr–Mn	3.97	2	7.7		
2	S <sub>2</sub>	Sr–Mn	3.56	3	9.6	0.48	0.19
		Sr-Mn	4.03	1	9.6		
1	$S_3$	Sr-Mn	3.42	2	8.9	0.56	0.23
		Sr–Mn	3.94	2	8.9		
1	$S_0$	Sr–Mn	3.47	2	6.8	0.23	0.10
		Sr–Mn	3.98	2	6.8		

R is the distance between Sr and Mn.  $S_0^2$ , an amplitude reduction factor (see SI Text, Eq 1), was set to 1.0.

The FT peak II in Sr EXAFS splits in the  $S_3$  state (Fig. 2B) suggesting changes in Sr-Mn distances. The best fit (Table 2) for the S<sub>3</sub> state is for two short and two long Sr-Mn distances. The Sr-Mn interaction at  $\approx 3.5$  Å shortens to  $\approx 3.4$  Å, and the distance at  $\approx 4.0$  Å decreases to  $\approx 3.9$  Å during the S<sub>2</sub>-to-S<sub>3</sub> transition. The fact that Mn-Sr interactions change during the S<sub>2</sub>-to-S<sub>3</sub> transition is also in agreement with the different efficiency of Ca depletion in the S<sub>3</sub> state from that in other S states (33). Ca/Sr-depleted PS II cannot advance to the S3 state; instead, a state designated S2YZ\* is formed in which the Mn4 core structure is close to that of the S2 state and does not resemble the structure of the native  $S_3$  state (12, 29). We propose that the single  $\mu_3$ -oxygen of the Mn<sub>4</sub>Ca structure is important for the formation of the S<sub>3</sub> state and that its properties are significantly altered in the absence of a coordinated Ca/Sr atom.

Completing the catalytic cycle in the  $S_3$ -to- $S_0$  transition, the  $di-\mu$ -oxo-bridged Mn–Mn interactions exhibit shortening (Table

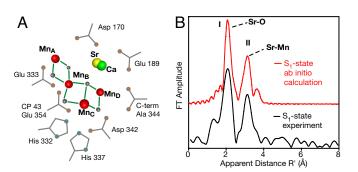


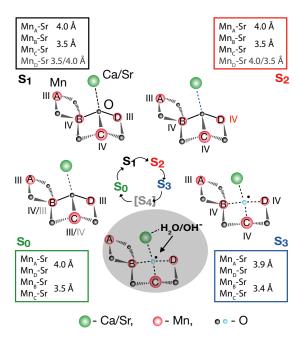
Fig. 3. (A) Ca atom (in green) and Sr atom (in yellow) were placed in the structural model derived from a polarized Mn EXAFS experiment on PS II single crystals (model II) (18) in the S<sub>1</sub> state. We note that the ligand/residues around the Mn<sub>4</sub>Ca cluster in this model are putative. The number of the Sr-Mn vectors and the Sr-Mn distances are from the results of fits to Sr EXAFS data (fit 1 in Table 2); (Sr–Mn<sub>B,C,D</sub> at  $\approx$ 3.5 Å and Sr–Mn<sub>A</sub> at  $\approx$ 4.0 Å). Only small changes in the Sr atom position are required, compared with the position of the Ca determined from polarized EXAFS studies of PS II crystals, to satisfy the Sr EXAFS data from Sr-PS II. (B) The Fourier transforms of the Sr EXAFS spectra from Sr-PS II of the S<sub>1</sub> state calculated by using the ab initio FEFF8 program for the placement of the Sr atom that is in accord with the best fit number 1 in Table 2. Coordinates of model II (18) with Sr as in fit 1 (Table 2) were used as input. B shows that peak II in the Sr EXAFS spectrum of PS II originates from Sr-Mn interaction. There is very good agreement between the experimental FTs from the Sr EXAFS spectra from Sr-PS II with the calculated FT for the model.

1) and recovery of peak III intensity in the Mn EXAFS (Fig. 1B). From Sr EXAFS, we can see that the Sr-Mn distances at 3.42 Å and at 3.94 Å increase to 3.47 Å and 3.98 Å, respectively. During the S<sub>0</sub>-to-S<sub>1</sub> transition, the Sr-Mn distances increase further to  $\approx$ 3.5 Å and  $\approx$ 4.0 Å.

The generally accepted proton release pattern during the  $S_0$ -to- $S_1$ ,  $S_1$ -to- $S_2$ ,  $S_2$ -to- $S_3$ , and  $S_3$ -to- $S_0$  transitions is 1, 0, 1, 2, respectively; although it has been reported that the proton release pattern is pH-dependent (34). EXAFS can provide only indirect information about protons, by the effect that protonations/deprotonations might have on the Mn-Mn/Sr(Ca) and Mn-O, N ligand atom distances. The shortening of one Mn-Mn distance observed during the S<sub>0</sub>-to-S<sub>1</sub> transition is compatible with the deprotonation of an hydroxo (OH<sup>-</sup>) bridge between Mn atoms to an oxo  $(O^{2-})$  bridge (35, 36), and consistent with the release of one proton during this transition. During the S<sub>1</sub>-to-S<sub>2</sub> transition there is no net release of protons and there are no significant changes in Mn–Mn/Ca distances. There is release of a proton during the S<sub>2</sub>-to-S<sub>3</sub> transition, and the absence of any shortening in Mn–Mn distances allows us to speculate that the proton release proceeds differently for this transition. There are no EXAFS studies of S states between S<sub>3</sub> and S<sub>0</sub>, when two protons are released. One speculation is that a proton is released from a hydroxo group that is a ligand of Ca/Mn (or from exogenous OH<sup>-</sup>/H<sub>2</sub>O) that is involved in the O-O bond formation, whereas another proton is released from a H<sub>2</sub>O ligand of Ca/Mn (or from exogenous H<sub>2</sub>O), which can be incorporated as a bridging OH<sup>-</sup> ligand in the S<sub>0</sub> state. Ca, which can accommodate seven or eight ligands, is ideally suited for ferrying in H<sub>2</sub>O/OH<sup>-</sup> groups to the catalytic site.

Fig. 4 summarizes the structural changes accompanying the S<sub>0</sub> to S<sub>3</sub> catalytic cycle transition as deduced from Mn and Sr EXAFS. On the oxidation of one Mn atom, probably Mn<sub>D</sub> (37, 38), during the  $S_1$ -to- $S_2$  transition, it is possible that the  $S_1$ -to- $S_2$  transition, it is possible that the  $S_1$ -to- $S_2$  transition is possible that the  $S_2$ -to- $S_2$ moves away from one Mn (Mn<sub>B</sub>, Mn<sub>C</sub>, or Mn<sub>D</sub>) leading to the change from 3:1 short:long Sr-Mn distances to a 2:2 ratio in the S<sub>2</sub> state. Alternatively, the 2:2 ratio can be present already in the S<sub>1</sub> state (lower fit quality; Table 1), thus remaining unchanged in the S<sub>1</sub>-to-S<sub>2</sub> transition. During the S<sub>2</sub>-to-S<sub>3</sub> transition, although the ratio of short-to-long vectors remains 2:2, there is a significant decrease in the Sr-Mn distances. From the Mn EXAFS the di- $\mu$ -oxo-bridged Mn–Mn distances increase during the S<sub>2</sub>-to-S<sub>3</sub> transition; from  $\approx$ 2.7–2.8 Å to  $\approx$ 2.8–2.9 Å (Table 1). This result is similar to that detected by Mn EXAFS using PS II preparations from spinach (29). Future studies should aim at

<sup>\*</sup>The number of Sr–Mn interactions was fixed at integer values of 0, 1, 2, 3, and 4.  $\sigma$  is the Debye–Waller factor.  $\Phi$  and  $arepsilon^2$  are the goodness-of-fit parameters. Details about parameters and fitting methodology are described in SI Text and SI Fig. 8.



Schematic of the structural changes accompanying the S state transitions in the Mn<sub>4</sub>Ca(Sr) cluster is placed within the context of the recent structural model II from single-crystal x-ray spectroscopy. The critical transition is the S<sub>2</sub>-to-S<sub>3</sub> advancement, when the Mn-Mn di-μ-oxo bridge distances of the Mn<sub>4</sub> core become elongated from ≈2.7–2.8 Å to ≈2.8–2.9 Å. Simultaneously, Sr is drawn closer to the Mn core with the Sr–Mn interaction at  $\approx 3.5$ Å shortening to  $\approx$ 3.4 Å and at  $\approx$ 4.0 Å distance decreasing to 3.94 Å. We propose that this significant change is triggered by the ligand-centered oxidation of the oxygen atom that bridges the Mn with the Ca atoms. The Ca(Sr)-Mn distances in all of the S states are indicated in the boxes next to the S states. The critical bridging oxygen atom is shown in blue. The dashed line (blue) between Ca and the bridging atom in the S2 state indicates the possible movement of Ca closer to two of the Mn atoms. The four dashed lines in the  $S_3$  state indicate a change in the structure around the  $\mu_3$ -oxo-bridged oxygen, probably from a tetrahedral to tetragonal distortion and leading to the changes in the Mn-Mn and Ca(Sr)-Mn distances. We note that a similar mechanism can be proposed by using model III (SI Fig. 9) from single-crystal EXAFS (18), that involves the oxidation of a different bridging oxygen atom during the S2-to-S3 transition.

identifying which particular Mn-Mn moiety in the model increases in distance.

**Mechanism of Photosynthetic Water Oxidation.** It is difficult to rationalize the changes in the Mn–Mn and Mn–Sr distances without significant involvement of the bridging O atoms. One hypothesis is that during the S<sub>2</sub>-to-S<sub>3</sub> transition the oxidation occurs predominantly at a bridging oxygen ligand, triggering the structural changes in the OEC. An oxyl radical intermediate has been proposed on the basis of density functional theory (DFT) calculations (39, 40), including on a bridging position between two manganese atoms (41). Preliminary resonant inelastic x-ray scattering (RIXS) data show that the charge density change during the S<sub>2</sub>-to-S<sub>3</sub> transition is much smaller than during the S<sub>1</sub>-to-S<sub>2</sub> transition, supporting the hypothesis that the oxidation is predominantly ligand-centered.

We think that the observed structural changes in the S<sub>2</sub>-to-S<sub>3</sub> transition are compatible with a primarily ligand-centered oxidation. The oxidized ligand in the S<sub>3</sub> state can interconvert between a bridging and a terminal ligand O atom (OH or H<sub>2</sub>O) (42) resulting in an oxygen isotope exchange. Changes in Mn–O–Mn vibrational frequencies, the EPR properties (7, 27), rate of water exchange (43) on Ca to Sr substitution, and data presented here indicate that the critical oxygen atom is part of the Mn–O–Ca/Sr bridging structure.

Although, there are many proposed mechanisms for the photosynthetic water oxidation reaction that include variations on how the O-O bond is formed, there are two important favored mechanisms for water oxidation by the : (i) nucleophilic attack on Mn(V)=O or Mn(IV)-O by a metal (possibly Ca)bound water molecule or hydroxide (9, 44); or (ii) reaction of a Mn-oxo unit with predominant radical character with an oxo/ hydroxo/water ligand or an exogenous H<sub>2</sub>O (29). Except for PS II there are few structurally defined catalysts competent to oxidize water at room temperature; Ru and Mn complexes provide examples (44, 45). A theoretical investigation of the Ru catalyst supported a Ru<sup>IV</sup>-O moiety to promote the watersplitting reaction (46). Theoretical analysis of the OEC indicates that the formation of a low-lying ligand-oxygen radical precursor state may be required for forming the O-O bond (39). To reach this state, a structural rearrangement is needed at the S<sub>2</sub>-to-S<sub>3</sub> transition. First, the changes in the Mn-Mn and Ca(Sr)-Mn distances require a model that involves a bridging oxygen atom (Fig. 4). This is probably the same oxygen where the oxidation occurs in the S<sub>3</sub> state, which triggers the formation of the O-O bond. We note, however, that the oxidation of a different bridging oxygen cannot be excluded. The bridging oxygen atom that is oxidized depends on whether we choose model II or III (18) for illustrating the general scheme of the mechanism. Second, the decrease in the Ca(Sr)-Mn distance during the S<sub>2</sub>-to-S<sub>3</sub> transition favors the further formation of the O–O bond between a Ca-bound water or hydroxide and a Mn-bound oxygen. These two observations lead us to propose that the Ca-bound water or hydroxide and a critical oxo-bridging atom with predominantly radical character are the oxygen atoms involved in the formation of the all-important O-O bond in the water oxidation reaction.

## **Materials and Methods**

Sample Preparation. Sr-containing PS II (Sr-PS II) was prepared as described in ref. 27, with the exception that the betaine concentration was decreased to 1 M. After elution from the Ni-affinity column, Sr-PS II was washed and concentrated by using centrifugal filtration devices (Ultrafree-15; Millipore). Sr-PS II was finally resuspended in 50% glycerol, 1 M betaine, 15 mM CaCl<sub>2</sub>, 15 mM MgCl<sub>2</sub>, 40 mM MES, pH 6.5 (adjusted with NaOH). Concentration of the Sr-PS II samples was  $\approx$ 15 mg of ChI/mI.

EXAFS sample holders with inner dimension of  $18 \times 2.5 \times 0.8$  mm were filled with Sr-PS II samples. After dark adaptation for 1 h at room temperature, a freshly prepared stock solution (50 mM in ethanol) of phenyl-p-benzoquinone (PPBQ) was added to obtain a final concentration of 500  $\mu$ M. PPBQ (Eastman Kodak) that was used in these experiments was recrystallized several times from ethanol. All of the flash illumination, EPR, and x-ray absorption measurements were performed directly on samples mounted in these holders. The samples were immediately frozen to 77 K in liquid nitrogen after flash laser illumination.

**Metal Quantitation.** Metal (Mn and Sr) quantitation was done by using a Perkin–Elmer 3110 Atomic Absorption Spectrometer equipped with an HGA 600 furnace. The quantitation yielded 37  $\pm$  1 Chl/4 Mn and 1.0  $\pm$  0.1 Sr/4 Mn.

Flash-Induced Illumination of PS II. A frequency-doubled (532 nm) Nd:YAG laser was used (8-ns pulsewidth) for flash illumination. To maintain maximal synchronization of the PS II centers on flash illumination, the fast recombination reaction between both the  $S_2$  and  $S_3$  states and the reduced form of the redox-active tyrosine residue  $Y_D$  must be suppressed. This was achieved by the application of one preflash, followed by a 60-min dark-adaptation period at room temperature. This procedure synchronizes the PS II centers into predominantly the  $S_1Y_D^{ox}$  state. Each sample was then given zero, one, two, three, four, five, or six flashes at room temperature, with 1.5-s intervals between individual flashes. The light was focused on the sample by using cylindrical lenses. After the last flash, the samples were frozen immediately (within 1 s) in liquid nitrogen. The EPR spectra were collected, and the samples were stored at 77 K for further use in the XAS experiments. Alternatively,  $S_2$  samples were prepared by 1-min continuous illumination at 200 K.

**Control with Inactive Sr-PSII.** The *inactive* Sr-PSII was prepared by adding 10  $\mu$ l of a stock solution (100 mM) of hydroxylamine (NH<sub>2</sub>OH) directly to the intact

Sr-PS II in the Lucite sample holder. After incubation for 20 min at room temperature in the dark, the sample was frozen in liquid nitrogen. NH2OHtreated samples are indicated as HYD in the figures.

EPR Spectroscopy. Low-temperature X-band EPR spectra were recorded by using a Varian E109 EPR spectrometer equipped with a model 102 microwave bridge. For the multiline-signal measurements, the sample temperature was maintained at 8 K with use of an Air Products LTR liquid helium cryostat. Spectrometer conditions were as follows: microwave frequency, 9.21 GHz; field modulation amplitude, 32 G at 100 kHz; microwave power, 30 mW. EPR multiline-signal amplitudes were quantified by adding peak-to-trough amplitudes of four of the hyperfine lines upfield from g = 2, as indicated in SI Fig. 5A. For each sample, the four designated S2 state multiline EPR signal peaks were quantified by using the amplitude of the  $Fe^{III}$  signal at g = 4.3 as an internal reference. The averaged results are shown as points in SI Fig. 5B normalized to the 1F value as 100%. The intensity of the multiline signal in the 1F samples and samples after continuous illumination were the same (within  $\pm$  10% uncertainty of the quantitative EPR technique), confirming the saturation of the sample with single laser flash illumination.

We used the Kok model as described in Messinger et al. (28) to calculate the S state population for each flash number and have compared the calculated S<sub>2</sub> state values (solid line, normalized to be 100% for 1F amplitude) to the normalized amplitudes (squares) shown in SI Fig. 5B (SI Tables 3 and 4). The error between the calculated and measured S2 state populations was minimized. because of factors such as redox equilibrium between the cofactors in PS II, it is inevitable that some dephasing occurs while the OEC is advanced through the various S states. The original Kok model explains this by assuming two parameters: the miss probability ( $\alpha$ ) accounts for the percentage of centers that do not advance in each flash, and the double-hit probability ( $\beta$ ) describes the percentage of centers that make two turnovers in a single flash. The value of  $\beta$  was first set to 0 based on the short pulsewidth of the Nd:YAG laser pulses. The possibility of double hits was considered in fit 2 and did not significantly improve the fit quality (SI Table 3). Oscillation characteristics obtained by more precise  $O_2$  evolution measurements or with UV-absorption changes give comparable miss probability (27).

**EXAFS Data Collection.** X-ray absorption spectra were collected at the Stanford Synchrotron Radiation Laboratory (SSRL) on beamline 9-3 at electron energy 3.0 GeV and an average current 70-100 mA. The intensity of the incident x-rays

- 1. Wydrzynski T, Satoh S (2005) Photosystem II: The Light-Driven Water: Plastoquinone Oxidoreductase (Springer, Dordrecht).
- 2. Peloquin JM, Britt RD (2001) Biochim Biophys Acta 1503:96-111.
- 3. Carrell TG, Tyryshkin AM, Dismukes GC (2002) J Biol Inorg Chem 7:2-22.
- 4. Hasegawa K, Ono TA, Inoue Y, Kusunoki M (1999) Chem Phys Lett 300:9-19.
- 5. Yachandra VK, Sauer K, Klein MP (1996) Chem Rev 96:2927-2950.
- Sauer K, Yano J, Yachandra VK (2005) Photosynth Res 85:73-86.
- 7. Chu HA, Hillier W, Law NA, Babcock GT (2001) Biochim Biophys Acta 1503:69-82.
- 8. Debus RJ, Strickler MA, Walker LM, Hillier W (2005) Biochemistry 44:1367–1374.
- 9. Ferreira KN, Iverson TM, Maghlaoui K, Barber J, Iwata S (2004) Science 303:1831–1838.
- 10. Loll B, Kern J, Saenger W, Zouni A, Biesiadka J (2005) Nature 438:1040-1044.
- 11. Latimer MJ, et al. (1995) Biochemistry 34:10898-10909.
- 12. Latimer MJ, DeRose VJ, Yachandra VK, Sauer K, Klein MP (1998) J Phys Chem B 102:8257-8265
- 13. Cinco RM, et al. (1998) J Phys Chem B 102:8248-8256.
- 14. Cinco RM, et al. (2002) Biochemistry 41:12928-12933.
- 15. Cinco RM, et al. (2004) Biochemistry 43:13271-13282.
- 16. Yano J, et al. (2005) J Am Chem Soc 127:14974-14975.
- 17. Pushkar Y, et al. (2006) J Biol Chem 282:7198-7208.
- 18. Yano J, et al. (2006) Science 314:821-825.
- 19. Boussac A, Rutherford AW (1988) Biochemistry 27:3476-3483.
- 20. Vrettos JS, Stone DA, Brudvig GW (2001) Biochemistry 40:7937-7945.
- 21. Westphal KL, Lydakis-Simantiris N, Cukier RI, Babcock GT (2000) Biochemistry 39:16220-16229.
- 22. Chu HA, Sackett H, Babcock GT (2000) Biochemistry 39:14371-14376.
- 23. Kimura Y, Hasegawa K, Ono T (2002) Biochemistry 41:5844-5853.

was monitored by a N2-filled ion chamber (I0) in front of the sample. The radiation was monochromatized by a Si (220) double-crystal monochromator. To reduce the sample damage by x-radiation, the incident x-ray beam was defocused at the sample position. The total photon flux on the sample was limited to 1  $\times$  10<sup>7</sup> photons per  $\mu$ m<sup>2</sup>, which was determined to be nondamaging on the basis of detailed radiation-damage studies of PS II solution samples (30). The samples were protected from the beam during spectrometer movements between different energy positions by a shutter synchronized with the scan program. The samples were kept at 9  $\pm$  1 K in a He atmosphere at ambient pressure by using an Oxford CF-1208 continuous-flow liquid He cryostat. Data were recorded as fluorescence excitation spectra by using a germanium 30-element energy-resolving detector (Canberra Electronics). For Mn XAS, energy was calibrated by the preedge peak of KMnO<sub>4</sub> (6,543.3 eV), which was placed between two  $N_2$ -filled ionization chambers ( $I_1$  and  $I_2$ ) after

Conditions for acquiring Sr EXAFS data on T. elongatus Sr-PS II samples closely resemble those for spinach solution and oriented samples and described in Cinco et al. (13, 15). The x-ray flux at 16-17 keV was  $2.4 \times 10^7$ photons per  $\mu$ m<sup>2</sup> of sample. For energy calibration, we simultaneously measured the absorption spectrum of solid strontium acetate, whose edge peak was assigned the value 16,120.0 eV (15). Spectra were collected with 3-eV steps in the preedge region (15,970-16,070 eV), 1-eV steps from 16,070 to 16,134 eV, and 0.075-Å<sup>-1</sup> steps from k = 2.0-13.5 Å<sup>-1</sup>. Sr EXAFS data were recorded during three x-ray beam times for S<sub>1</sub>, S<sub>2</sub>, and S<sub>3</sub> samples and during two beam times for S<sub>0</sub> samples. PS II samples from different preparations were used for each of these experiments. Identical spectra were obtained for all of these different PS II preparations.

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- 24. Kimura Y, Hasegawa K, Yamanari T, Ono TA (2005) Photosynth Res 84:245-250.
- 25. Strickler MA, Walker LM, Hillier W, Debus RJ (2005) Biochemistry 44:8571-8577.
- 26. Suzuki H, Taguchi Y, Sugiura M, Boussac A, Noguchi T (2006) Biochemistry 45:13454-13464.
- 27. Boussac A. et al. (2004) J Biol Chem 279:22809-22819.
- 28. Messinger J, et al. (2001) J Am Chem Soc 123:7804-7820.
- 29. Liang W, et al. (2000) J Am Chem Soc 122:3399-3412.
- 30. Yano J, et al. (2005) Proc Natl Acad Sci USA 102:12047-12052.
- 31. Mishra A, et al. (2007) Chem Commun 15:1538-1540.
- 32. Kuntzleman T, McCarrick R, Penner-Hahn J, Yocum C (2004) Phys Chem Chem Phys 6.4897-4904
- 33. Boussac A, Rutherford AW (1988) FEBS Lett 236:432-436.
- 34. Rappaport F, Lavergne J (2001) Biochim Biophys Acta 1503:246-259.
- 35. Baldwin MJ, et al. (1994) J Am Chem Soc 116:11349-11356.
- 36. Robblee JH. et al. (2002) J Am Chem Soc 124:7459-7471.
- 37. Chu HA, Hillier W, Debus RJ (2004) Biochemistry 43:3152-3166.
- 38. Kimura Y, Mizusawa N, Yamanari T, Ishii A, Ono T (2005) J Biol Chem 280:2078-2083.
- 39. Siegbahn PEM, Crabtree RH (1999) J Am Chem Soc 121:117-127.
- 40. Lundberg M, Blomberg MRA, Siegbahn PEM (2004) Inorg Chem 43:264-274.
- 41. Siegbahn PEM (2000) Inorg Chem 39:2923-2935.
- 42. McEvoy JP, Brudvig GW (2006) Chem Rev 106:4455-4483.
- 43. Hendry G, Wydrzynski T (2003) Biochemistry 42:6209-6217.
- 44. Vrettos JS, Limburg J, Brudvig GW (2001) Biochim Biophys Acta 1503:229-245. 45. Yamada H, Siems WF, Koike T, Hurst JK (2004) J Am Chem Soc 126:9786-9795.
- 46. Yang X, Baik M-H (2006) J Am Chem Soc 128:7476-7485.